

Proximate Composition and Cholesterol Profile of Table Eggs from Hens Fed Different Proprietary Feeds in Ibadan, Nigeria

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Abstract.

Proprietary feed labels are by regulation tailored to the requirements of the target animal. However, different proprietors lay claim to equal efficacy of feed whilst there could be variations in the actual composition and claims on the labels. This study was aimed at investigating the effects of four proprietary commercial layer feeds in Ibadan, Nigeria on the proximate composition and cholesterol profile of laid eggs. Black Bovan Nera hens (n=96) aged 24-week, were randomly allotted to four different commercial feed A, B, C and D, respectively. Each treatment was in triplicates of eight birds. Hens were managed under a cage system with experimental diets and water provided ad-libitum. At day 56 of feeding the hens on experimental feeds, eggs were collected for proximate and cholesterol profile determinations. Feed type had significant effects (P<0.05) on total cholesterol and low density lipoproteins (LDL). Total cholesterol was higher (P<0.05) in eggs from hens on Feed-A (1195.98mg/dL) and lowest in those from Feed-C (685.07mg/dL). Also, higher (P<0.05) LDL was in eggs of hens on Feed-A (971.55 mg/dL) and least in those on Feed-C (173.71 mg/dL). There was no effect of feeding the different commercial feeds to hens on high density lipoprotein, triglycerides, very low density lipoprotein, metabolizable energy and proximate composition. True protein was however, higher (P<0.05) in eggs of hens on Feed-A (6.29%) than those on other feeds. Thus, nutrients composition and choice of feed for laying hens may greatly influence true protein, total cholesterol and LDL content of eggs.

Keywords: Cholesterol, Crude protein, Lipoprotein, Triglyceride, True protein

Introduction

Eggs constitute an important part of human diet because of its high quality protein (Forson *et al.*, 2011). Egg, a source of complete protein with good quality amino acid profile is one of the most frequently consumed foods in the world, with chicken egg being the highest consumed in western societies (Muller and Tobin, 1996). Keeping chickens for egg production has become one of the fastest ways of meeting

the protein demands in a nation's population in that, no taboo or religion forbids its consumption. Also, eggs could be consumed absolutely when fresh without any need for refrigeration and storing the left-over (Farell, 2013; Ogunwole *et al.*, 2015a).

The nutrition of birds has direct effect on the quality of eggs (Cherian *et al.*, 2002). Insufficient levels of nutrients in hen diet could impair the efficiency of

production, and eggs laid could be of inferior quality. Reports have shown that cholesterol and proximate composition of egg are affected by various factors such as breed, management and nutrition (Clum, 1996; Fakai *et al.*, 2015).

Developing countries have been reported to have reduced intake of eggs, a cheap animal source of balanced amino acids. The ascribed reason was that cholesterol and saturated fats especially from animal sources are bad (Rahimi, 2005; Mohsin, 2013). Whereas, polyunsaturated fatty acids which are abundant in animal products like egg, has been reported to reduce the chance of atherosclerosis and stroke (Lada and Rudel, 2003). Rahimi (2005) and Farell (2013) reported that most people, particularly, in developing countries believed in dangers of eating eggs due to erstwhile instinctive hype on saturated fats and cholesterol consumption. In the developing countries, diets are mainly plant-based with low cholesterol, except for the few affluent (FAO, 2007). Based on recent comprehensive study on cholesterol from more than a million patients, there was no evidence to prove that having high levels of cholesterol causes heart disease (Linnekin, 2018).

Nutritional strategies or dietary manipulations to reduce cholesterol concentration as well as to improve egg protein content has been advocated (NRC, 1994; Jurgens, 2002). Quality commercial feeds with appropriate nutritional values capable of achieving efficient performance and production without losing sight of the ensuing effects on functional composition of eggs is highly imperative in layers. This is in a quest to ensure that the health or nutrition concerns of egg consumers are properly taken into consideration. There is the need to investigate the effect of feeding

different proprietary feeds to layers on the composition of eggs they produce.

Few reports have documented the effects of feeding different proprietary feeds on chemical composition of eggs in Nigeria. Earlier documentations were on supplemental effects of the different proprietary vitamin-mineral premixes under two rearing systems on the proximate composition (Yan *et al.*, 2014; Ogunwole *et al.*, 2015a) and the lipids profile of eggs (Van *et al.*, 2004; Ogunwole *et al.*, 2015b). The present study was therefore designed to evaluate the effect of feeding four proprietary commercial feed to laying hens on yolk cholesterol, proximate and true protein composition of eggs produced by commercial pullets.

Materials and Methods

Experimental site

The experiment was conducted at the Poultry Unit of the Teaching and Research Farm, University of Ibadan, Ibadan. The farm is situated in the derived savanna vegetation belt of Nigeria. The study area lies between longitude 7°27.05 north and 3°53.74 of the Greenwich Meridian east at an altitude 200m above sea level. Average temperature and relative humidity of the location is between 23-42 °C and 60-80%, respectively (SMUI, 2018).

Experimental birds and management

A total of 96 Bovan Nera Black laying hens aged 24 weeks were randomly allotted to four treatments each in triplicate of eight birds per replicate in a completely randomized design. The hens were managed under battery system. The hens were offered their respective proprietary diets Feed-A, Feed-B, Feed-C and Feed-D all through with water *ad libitum* for eight weeks (56 days) of feeding trial.

Experimental diets

Four different proprietary (commercial) feed were used for the experiment. The four commercial feeds were purchased in Ibadan, Oyo state Nigeria. The feeds were tagged Feed-A, B, C and D, respectively. Feed-A had metabolisable energy and crude protein of 2700 Kcal/kg and 15.00%, respectively while B, C and D had 2500 Kcal/kg and 16.50%; 2500 Kcal/kg and 16.00%; 2500 Kcal/kg and 16.50% metabolisable energy and crude protein, respectively. Details of the gross composition of the commercial feeds as shown in the attached nutrients label of the Feed is shown in Table 1.

Table 1: Gross composition of commercial feed as shown in the attached nutrients label

| Parameters | Feed Type | | | |
|-------------------|-----------|-------|-------|-------|
| | A | B | C | D |
| ME (Kcal/kg) | 2700 | 2500 | 2500 | 2500 |
| Crude protein (%) | 15.00 | 16.50 | 16.00 | 16.50 |
| Fat (%) | 5.00 | 5.00 | 5.00 | 5.00 |
| Crude fibre (%) | 10.00 | 6.00 | 6.00 | 6.50 |
| Calcium (mg) | 3.50 | 3.60 | 3.50 | 3.50 |
| Potassium (mg) | 0.40 | 0.45 | 0.45 | 0.45 |
| Lysine | 0.75 | 0.80 | 0.80 | 0.80 |
| Methionine | 0.30 | 0.34 | 0.34 | 0.34 |
| Salt (%) | 0.30 | 0.30 | 0.30 | 0.30 |

Determination of cholesterol profile

On day 56, two eggs per replicate were randomly sampled for analyses. The total cholesterol (TC), Triglycerides, High Density Lipoprotein (HDL), Low Density Lipoprotein (LDL) and Very Low Density Lipoprotein (VLDL) were analyzed according to AOAC (2005).

Proximate and true protein composition

Two eggs were randomly sampled from each replicate. They were broken, homogenized and analyzed for proximate composition (AOAC, 2000).Metabolizable energy (kcal/kg) of the different feed samples was estimated according to the procedure of Ponzenga (1985). The true

protein was determined by subtracting non protein nitrogen from the total nitrogen value of the diet and the residual nitrogen value multiplied by a factor of 6.38 (Sinaga et al., 2015)

Statistical analysis

Data were subjected to analysis of variance using the general linear procedure (SAS, 1999). The means were separated using Duncan's Multiple Range Test of the same software at $\alpha_{0.05}$.

Results

The metabolizable energy estimates and lipid profile of egg from hens on different proprietary feeds are shown in Table 2. There were no significant differences ($P>0.05$) in the metabolizable energy, high density lipoprotein, triglyceride and very low density lipoprotein contents of eggs from birds, fed the various proprietary diets. Metabolisable energy (ME kcal/kg) however ranged from 4056.32 (B) to 4110.79 (A), HDL from 192.45 (B) to 261.11 (A), triglyceride from 1204.80 (C) to 1483.20 (A) and VLDL from 240.96 (C) to 1483.20 (A). However, there were significant differences ($P<0.05$) in TC content of the eggs. The TC of eggs from hens onFeed-A (1195.98 mg/dL) was similar to Feed-B (941.55) but significantly higher ($P<0.05$) than those fromFeed-C (685.07 mg/dL) and Feed-D (722.59 mg/dL). The TC of eggs from hens on Feed-B, C and D however were similar ($P>0.05$). Significant differences ($P<0.05$) were also observed in the LDL content among treatments with eggs from hens on Feed-A (971.55 mg/dL) having significantly higher ($P<0.05$) LDL compared with those on Feed-B (454.551 mg/dL), C (173.71 mg/dL) and D (290.53 mg/dL).

Table 2: Metabolizable energy and lipid profile of egg yolk from hens fed four different proprietary feeds in Ibadan, Nigeria

| Parameters | Feed Type | | | | SEM |
|--------------------------------------|----------------------|----------------------|---------------------|---------------------|----------------------|
| | A | B | C | D | |
| ME (kcal/kg) | 4110.79 | 4056.32 | 4108.58 | 4090.31 | 13.72 ^{NS} |
| Total cholesterol (mg/dL) | 1195.98 ^a | 941.55 ^{ab} | 685.07 ^b | 722.59 ^b | 130.51 |
| Low density lipoprotein (mg/dL) | 971.55 ^a | 454.44 ^b | 173.71 ^b | 290.53 ^b | 153.82 |
| High density lipoprotein (mg/dL) | 261.11 | 192.45 | 270.39 | 253.78 | 89.85 ^{NS} |
| Triglyceride (mg/dL) | 1483.20 | 1473.28 | 1204.80 | 1233.60 | 121.78 ^{NS} |
| Very Low density lipoprotein (mg/dL) | 296.64 | 294.66 | 240.96 | 246.72 | 24.36 ^{NS} |

Means with different superscripts along same row are significantly different (P<0.05) –Where, A = Hens fed commercial feed type A; B = Hens fed commercial feed type B; C = Hens fed commercial feed type C; D = Hens fed commercial feed type 4. ME= metabolizable energy; NS = Not significant (P>0.05)

The proximate composition of eggs (g/100g) from hens fed four different proprietary feed in Ibadan, Nigeria are shown in Table 3. There were no significant differences (P>0.05) in the proximate composition of eggs from hens on different commercial feeds. Moisture content ranged from 73.32 in Feed-B to 75.89% in Feed-C, crude protein from 10.14 (C) to 11.39%

(B), ether extract from 13.09 (C) to 14.08% (B), ash 0.79 (A and D) to 1.18% (B) while NFE were 0.04, 0.03, 0.07 and 1.01% in Feed-A, B, C and D, respectively. True protein in Feed-A (6.29%) was similar to 5.24% in Feed D-4 but significantly higher than (P<0.05) 4.30 and 4.64% in Feed-B and C, respectively.

Table 3: Chemical composition of eggs fed four different proprietary feeds in Ibadan, Nigeria

| Parameters (%) | A | B | C | D | SEM |
|------------------------|-------------------|-------------------|-------------------|--------------------|--------------------|
| Moisture content | 74.95 | 73.32 | 75.89 | 74.73 | 0.22 ^{NS} |
| Crude protein | 10.91 | 11.39 | 10.14 | 10.23 | 0.18 ^{NS} |
| True protein | 6.29 ^a | 4.30 ^b | 4.64 ^b | 5.24 ^{ab} | 0.21 |
| Ether extract | 13.31 | 14.08 | 13.09 | 13.24 | 0.54 ^{NS} |
| Ash | 0.79 | 1.18 | 0.81 | 0.79 | 0.09 ^{NS} |
| Nitrogen free extracts | 0.04 | 0.03 | 0.07 | 1.01 | 0.02 ^{NS} |

Mean values with different superscript in a row differ significantly (P<0.05)NFE: Nitrogen Free Extract;Where, 1 = Hens fed commercial feed type A, 2 = Hens fed commercial feed type B, 3 = Hens fed commercial feed type C, 4 = Hens fed commercial feed type 4; NS = Not significant (P>0.05)

Discussion

Cholesterol are naturally found in the cell walls structure and required in the production of the steroid hormone vitamin D and bile salts which helps digestion (Fakai *et al.*, 2015). The differences in the cholesterol profile of eggs could be attributed to the varying energy content of the feeds. Feed-A had relatively higher energy content of 2700 kcal/kg compared with B, C and D. Vargas and Naber (1984), earlier correlated yolk cholesterol content with dietary energy balance and asserted that excessive energy intake, beyond maintenance and production requirements, increased body weight and cholesterol synthesis. Therefore, reserve cholesterol would be transferred to the egg yolk since layers releases its excess cholesterol in the eggs. Conversely, Fennema (1993) reported that variations in total yolk lipid content are more influenced by bird genetics than diet. Differences observed in cholesterol concentration in the present study could be attributed to diet as the evaluation was undertaken with birds of the same genetic strain and age.

Low density lipoprotein are erroneously referred to as 'bad cholesterol' because of their perceived potential to block blood vessels, preventing the flow of blood and thus elicit various cardiovascular diseases (Howard *et al.*, 2000; Tall *et al.*, 2001). The higher level of yolk LDL produced by hens on Feed-A may be attributed to the reduced level of crude protein and higher level of energy compared to others. The higher LDL recorded in eggs of hens on Feed-A despite the higher fibre levels contradicts the submission of Biswas *et al.* (2011) that increased dietary fibre would reduce LDL. Increased HDL was an indication of good healthy condition (Gordon *et al.*, 1989),

hygienic condition of the egg (Zemková *et al.*, 2007) and freedom from contaminants that might lead to increased peroxidation and deterioration (Blokhuis *et al.*, 2007).

There was no observable difference in metabolisable energy of eggs obtained from hens on the different treatments. Similar results were also obtained by Costa *et al.* (2004), Wu *et al.* (2007), Jalal *et al.* (2007) and Costa *et al.* (2009) in layers fed different ME levels. The authors inferred that dietary energy levels did not have any significant effect ($P>0.05$) on egg production. However, Araújo and Peixoto (2005) observed a reduced egg production ($P<0.05$) as dietary energy levels increased, while Valkonen *et al.* (2008) obtained increased egg production with higher dietary energy levels. These contentions may be explained by the fact that the energy levels higher than those recommended did not increase production, while energy deficiency decreased production. Leeson and Summers (2000) showed that increased energy intake had positive significant effects on egg weight.

The insignificant variation in proximate composition of eggs in this study conforms to report of Nys (2004) that egg displays very consistent compositions with regard to its content of total proteins, essential amino acids, total lipids, phospholipids, phosphorus and iron. Proteins are essential component of living cell; they are polymers of amino acid and the nutrient needed by human body for growth and maintenance of body cells (Fakai *et al.*, 2015). Egg provides means through which the protein needs of the populace could be met. Eggs has various use and contain many essential nutrients as it support life during embryonic growth and one of the nutritious and complete food known to man (Scott and

Ross, 2001). Gilbert (1979) reported that chicken egg generally contain about 12% by weight of protein. The result observed in this study however, was relatively lower compared to this but higher than values reported for Shika Brown (Fakai *et al.*, 2015)

The moisture content of a given sample simply refers to the water content of that sample (Fakai *et al.*, 2015). The amount of water in a food varies from low to high moisture levels in food (Elvan *et al.*, 2008). Fresh eggs were used in this analysis, and as such contained high amount of moisture. Moisture level in chicken eggs compared favourably to that obtained for duck (Fakai *et al.*, 2015) and laying chickens (Ogunwole *et al.*, 2015a, b). Hassan *et al.* (2008) stated that moisture contents of food above 15% would favour microbial activities which will result to food spoilage. Thus, moisture content of all the eggs exceeded 15% which indicated that eggs are highly perishable.

Lipids stored in the tissue are mobilized as a source of energy during stressful conditions, or during food deprivation (Gordon, 2002). The ether extracts ranged between 13.09 to 14.08%. The values obtained from this study were higher than those reported in literature for *Gallus domesticus*, *Numida melleagris* and *Columbia livia* (Adenowo *et al.*, 1999, Emmanuel *et al.*, 2011 and Fakai *et al.*, 2015) but comparable to that for local poultry eggs.

Ash represents the amount of mineral in a given sample (Fakai *et al.*, 2015). Samples with high concentration of various mineral elements were expected to speed up

metabolic processes, improve growth and development (Muhammad, 2011). Ash contents of eggs obtained from hens on Feed-B were similar to reported 0.94% ash of eggs (Gordon, 2002). Values of NFE obtained from this study were lower than those earlier reported for eggs (Ogunwole *et al.*, 2015a) while high true protein of eggs from hens on Feed-A could be attributed to higher fibre content of the feed that would reduce passage time and increased time for absorption of nutrients during digestion in the gut of hens.

Conclusion

Feeding different proprietary feeds to laying chickens had varying effects on the total cholesterol and low density lipoprotein compositions of eggs. There was no observable effect of feeding the different commercial feed on proximate composition of eggs. However, the true protein composition of eggs varied with the type of feeds given to hens in this study. Therefore, adequate cautions must be exercised in the choice of any feed for laying chickens by Poultry Nutritionists.

Conflict of Interest

Authors hereby declare that this study was undertaken in best scientific tradition devoid of any conflict or special interest in any particular product. In line with this avowal, further details of the test feeds are obtainable on enquiry from the corresponding author.

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References

- Adenowo, J. A., Awe, F. A., Adebambo, O. A and Ikeobi, C. N. (1999). Species variation in chemical composition of local poultry eggs. *Book of Proceeding: 26th Annual NSAP Conference 21-25 March*, University of Ilorin, Ilorin: 278-280.
- Araújo, J. S. and Peixoto, R. R. (2005). Níveis de energia metabolizável em ração para poedeiras de ovos marrons nas condições de inverno no extremo sul do Brasil. *Archivos de Zootecnia* 54: 13-23.
- Association of Official Analytical Chemists (2000) Official methods of analysis, 15th edition, Washington, DC.
- Association of Official Analytical Chemists (2005) Official method of Analysis. 18th Edition, Washington DC.
- Biswas, A. K., Kumar, V., Bhosle, S., Sahoo, J and Chatli, M. K. (2011). Dietary fibers as functional ingredients in meat products and their role in human health. *International Journal of Livestock Production* 2 (4):45-54.
- Blokhuis, H.J., Van Niekerk, T.F., Bessei, W., Elson, A., Guemene, D., Kjaer, J.B., Levrino, G.A.M., Nicol, C.J., Tauson, R., Weeks, C.A. and De Weerd, H.A.V. (2007). The LayWel project: welfare implications of changes in production systems for laying hens. *World Poultry Science Journal* 63: 101-114.
- Bolu, S.A. (2013). Vitamins in Poultry Nutrition. T-Babs printers. ISBN: 978-8113-02-8
- Cherian, G., Holsonbake, T.B. and Goeger, M.P. (2002). Fatty acid composition and egg components of specialty eggs. *Poultry Science* 81: 30-33.
- Chowdhury, S.R., Chowdhury, S.D. and Smith, T.K. (2002): Effects of dietary garlic on cholesterol metabolism in laying hens. *Poultry Science* 81: 1856-1861.
- Clum, N. J., Fritpatrick, M. P. and Dierenfeld, E. S. (1996). Effect of Diet on the nutritional content of whole prey. *Zoo Biology Journal* 15:525-537.
- Costa, F. G. P., Souza, H. C., Gomes, C. A. V., Barros, L. R., Brandão, P. A., Nascimento, G. A. J., Santos, A. W. R., Amarante Junior, V. S. (2004). Níveis de proteína bruta e energia metabolizável na produção e qualidade dos ovos de poedeiras da linhagem Lohmann Brown. *Ciência e Agrotecnologia* 28(6):1421-1427.
- Costa, F. G. P., Quirino, B. J. S., Givisiez, P. E. N. (2009). Poedeiras alimentadas com diferentes níveis de energia e óleo de soja na ração. *Archivos de Zootecnia* 58 (223): 405-41.
- Elkin, R. (2006). Reducing shell egg cholesterol content. I. Overview, genetic approaches, and nutritional strategies. *World's Poultry Science Journal* 62: 665-687.
- Elvan, C. E., Simmonds, N and MCollin, E. V. (2008). The newer knowledge of nutrition. 4th Edition Macmillian Co. New York. Pp 873-890.
- Emmanuel, T. E., Omole, J. O., Joseph, E. and Utu, B. A. (2011). Variation in micronutrients contents and lipid profile of some avian eggs. *America Journal of Experimental Agriculture*, 1(4): 343-354.
- Fakai, I. A., Sani I. and Olalekan, O. S. (2015). Proximate composition and cholesterol content of egg obtained from various bird species. *Journal of Harmonized Research in Medical and Health Science*. 2(2), 18-25 ISSN 2395-6046
- FAO (2007). The state of food and agriculture. Paying farmers for environmental services. Food and agriculture organization of united nations, Rome.
- Farrell, D. (2013). How important is cholesterol in eggs? The role of poultry in human nutrition. Poultry Development Review. Retrieved from <http://www.fao.org/docrep/019/i3531e/i3531e02.pdf>
- Fennema, O. R. (1993). *Química de los alimentos*. Acribia. Zaragoza. Tr. de la 2a. ed. original en inglés: Food chemistry, Ed. Marcel Dekker Pp 1-18.
- Forson, A., Axivor, J. E., Banini, G. K., Nuviadenu, C. and Debrah, S. K. (2011). Evaluation of some elemental variation in raw egg yolk and

- egg white of domestic chicken, guinea, fowl and duck eggs. *Annals of Biological Research*, 2: 676-680.
- Gilbert, A. B. (1979). Form and Function in Birds (King A.S., MacLelland J. (Eds.), Academic Press, Edinburgh, Pp 237-360.
- Gordon, D.J., Probstfield, J.L., Garrison, R.J., Neaton, J.D., Castelli, P., Knoke, J.D., Jacobs D., Bangdiwala, S., Tyroler, H.A. (1989). High density lipoprotein cholesterol and cardiovascular disease for prospective American studies. *Circulation* 79(1): 8-15.
- Gordon, M. W. (2002). Vitamin and mineral in contemporary nutrition. 5th Edition. M.cGram Hill New York. Pp. 239-281.
- Hargis, P. S. (1988): Modifying egg yolk cholesterol in domestic fowl - A review. *WorldsPoultry Science Journal* 44:17-19.
- Hassan, L., Dangogo, S. M, Umar, K. J. Saidu, I. and Folorunsho F. A. (2008). Proximate, mineral and antinutritional factors of *Daniellicioliveri* seed kernel, *Chemical and Classical Journal* 5: 31-36.
- Howard, B. V., D. C. Robbins, M. L. Sievers, E. T. Lee, D. Rhoades, R. B. Devereux, L. D. Cowan, R. S. Gray, T. K. Welty, O. T. Go, Wm. J. Howard (2000) LDL cholesterol as a strong predictor of coronary heart disease in diabetic individuals with insulinresistance and low LDL the strong heart study. *Arteriosclerosis, Thrombosis, and Vascular Biology* 20:830-835. doi: 10.1161/01.ATV.20.3.830
- Jalal, M. A., Scheideler, S. E. and Pierson, E. M. (2007). Strain response of laying hens to varying dietary energy levels with and without avizyme supplementation. *Journal of Applied Poultry Research*, 16: 289-295,
- Jurgens, M. H. (2002). Animal feeding and Nutrition. 9th Ed. Kendall/Hunt Pub. Coy. Iowa, USA. Pp. 146.
- Kanchana, G. and Jeyanthi, G. P. (2010). The Effect of Supplementation of diet with Vitamin-E and feeding on n-3 fatty acid and alpha-tocopherol content and oxidative stability of eggs. *Animal Feed Science Technology Journal* 9 (72): 33-40.
- Krawczyk, J. and Gornowicz, S. (2009). Effect of layer age and egg production level on changes in quality traits of eggs from hens of conservation breeds and commercial hybrids. *Annals of Animal Science Journal*, 9(2): 185-193.
- Lada A. T. and Rudel L. L. (2003). Dietary monounsaturated versus polyunsaturated fatty acids: Which is really better for protection from coronary heart disease? *Current Opinion in Lipidology* 14: 41-46.
- Leeson, S., Summers, J. D. (2000). Broiler breeder production. University books, Guelph, Ontario. Pp. 329.
- Linnekin, B. (2018). New Research Confirms We Got Cholesterol All Wrong. <https://reason.com/archives/2018/09/22/new-research-confirms-we-got-cholesterol>
- Marks, H. L. and Washburn, K. W. (1977). Divergent selection for yolk cholesterol in laying hens. *British Poultry Science*, 18 (2):179-188.
- Mohiti-Asli, M. and Zaghari, M. (2010). Effect of dietary vitamin E or C on egg yolk cholesterol. *Biology of Trace Element Research Journal* 138 (1-3):60-68.
- Mohsin Ali Baloch (2013). Serum Lipid Profile Hypertensive Patients in Hyderabad, Pakistan. *Nature and Science*. 11 (9):115-118. ISSN:1545-0740. <http://www.sciencepub.net/nature>
- Muhammad, M. A. (2011). Nutritional and antinutritional analysis of gardensa aqualice. M.Sc. Dissertation submitted to Post Graduate School Usman Danfodio University Sokoto, Pp88.
- Muller, H. G., and Tobin, G. (1996). Nutrition y ciencia de los alimentos, Zaragoza: Editorial Acribia.
- NRC.(National Research Council) (1994). Nutrient Requirements of Poultry: Ninth Revised Edition. National Academy Press, Washington D.C

- Nys, Y., Gautron, J., Garcia-Ruiz, J. M., Hinche, M. T. (2004). Avian eggshell mineralization: biochemical and functional characterization of matrix proteins. *Comptes Rendus palevol* 3: 549-562.
- Ogunwole, O. A., Ojelade, A. Y. P., Oyewo, M. O. and Essien, E. A. (2015a). Proximate composition and physical characteristics of eggs from laying chickens fed different proprietary vitamin-mineral premix under two rearing systems during storage. *International Journal of Food Science and Nutrition Engineering* p-ISSN: 2166- 5168 e-ISSN: 2166-51922015; 5(1): 59-67
doi:10.5923/j.food.20150501.08.
- Ogunwole, O. A., Ojelade, A. Y. P., Essien, E. A. and Oyewo, M. O. (2015b) Lipid Profile of Eggs from Laying Chickens Fed Five Proprietary Vitamin-Mineral Premixes under Two Rearing Systems as Influenced by Duration of Storage. *Food and Public Health*, 5(1): 10-16. DOI: 10.5923/j.fph.20150501.02
- Pauzenga. 1985. Feeding parent stock. *Zoo Technical International*. Pp 22-23.
- Rahimi, G. (2005). Dietary Forage Legume (*Onobrychis altissima grossh.*) Supplementation on Serum/Yolk Cholesterol, Triglycerides and Egg Shell Characteristics in Laying Hens. *International Journal of Poultry Science* 4 (10): 772-776.
- SAS (1999). SAS/STAT User's Guide. Version 8 for windows. SAS Institute Inc., SAS Campus Drive, Cary, North Carolina, USA.
- Scott, T. A., Silversides, F. G and Ross, D. A. (2001). Effect of storage and layer age on quality of eggs from two lines of hens. *Poultry Science* 80: 1245-1248.
- Sinaga, S. M., L. Margata and J. Silalahi. (2015). Analysis of Total Protein and Non Protein Nitrogen in Coconut Water and Meat (*Cocos Nucifera* L.) by using Kjeldahl Method Analysis. *International Journal of PharmTech Research*, 8 (4): 551-557
- Zemková, E., J. Simeonovová, M. Lichovníková, and K. Somerlíková. (2007). The effects of housing systems and age of hens on the weight and cholesterol concentration of the egg, *Czech Journal of Animal Science*, 52, (4): 110–115
- SMUI. (2018). Satellite Map of University of Ibadan, <https://latitude.to/articles-by-country/ng/nigeria/15223/university-of-ibadan>
- Tall, A. R., Wang, N., and Mucksavage, P. (2001). Is it time to modify the reverse cholesterol transport model? *Journal of Clinical Investigation*, 108:1273–1275.
- Valkonen, E., Venalainen, E.; Rossow, L. (2008). Effects of dietary energy content on the performance of laying hens in furnished and conventional cages. *Poultry Science Journal*, 87: 844-852.
- Van Den Bran, H., Parmentier, H. K., Kemp, B. (2004). Effects of housing systems (outdoor Vs cages) and age of laying hen on egg characteristics. *British Poultry Science*, 45, 745-752.
- Vargas, R. and Naber, E. (1984). Relationship between dietary fiber and nutrient density and its effects on energy balance, egg yolk cholesterol and hen performance. *Journal of Nutrition*, 114 (4):645-652.
- Wu, G., Bryant, M. M., Gunawardana, P. (2007) Effect of nutrient density on performance, egg components, egg solids, egg quality, and profits in eight commercial Leghorn strain during phase one. *Poultry Science*, 86: 691-697.
- Yan, F. F. Hester, P. Y. and H. W. Cheng. (2014). The effect of perch access during pullet rearing and egg laying on physiological measures of stress in White Leghorns at 71 weeks of age. *Poultry Science* 93:1318–1326
<http://dx.doi.org/10.3382/ps.2013-03572>